Wilke, M., Appel, K., Vincze, L., Schmidt, C., Borchert, M., Pascarelli, S. (2010): A confocal set-up for micro-XRF and XAFS experiments using diamond-anvil cells. - Journal of Synchrotron Radiation, 17, 669-675

DOI: 10.1107/S0909049510023654

A confocal setup for micro-XRF and XAFS experiments using diamond anvil cells

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revised version June 2010

Synopsis

A confocal setup for micro-XRF and XAFS for in-situ high-pressure and temperature experiments is presented.

Abstract

A confocal setup is presented that improves micro-XRF and XAFS experiments with high-pressure diamond anvil cells (DAC). In this experiment, a probing volume is defined by the focus of the incoming synchrotron radiation beam and that of a polycapillary X-ray half-lens with a very long working distance, which is placed in front of the fluorescence detector. This setup enhances the quality of the fluorescence and XAFS spectra, and thus the sensitivity for detecting elements at low concentrations. It efficiently suppresses signal from outside the sample chamber, which stems from elastic and inelastic scattering of the incoming beam by the diamond anvils as well as from excitation of fluorescence from the body of the DAC.

Keywords: XRF, XAFS, high-pressure, aqueous fluids

Introduction

The combination of high-pressure devices and X-ray techniques based on synchrotron radiation has become a standard experimental setup to study materials at high pressure and temperature. A large number of these studies is performed using diamond anvil cells (DAC) due to the low absorption of hard X-rays by diamond and due to the extreme pressure conditions that can be reached (e.g. Keppler & Frost 2005, Boehler 2005). While X-ray diffraction using these cells has been a standard technique for quite a long time, the application of X-ray fluorescence (XRF) analysis to quantify element concentrations of the sample in the DAC has only been applied lately (Schmidt & Rickers 2003, Sanchez-Valle et al. 2003). These and subsequent studies (Sanchez-Valle et al. 2004, Schmidt et al. 2006, Schmidt et al. 2007, Manning et al. 2008, Borchert et al. 2009) have shown that detection limits in the lower ppm-range can be reached for elements down to atomic number 22 (Ti). These high sensitivities have been achieved using an intense focused synchrotron radiation beam and/or modifications of the cells to optimize the signal-to-background ratio of the fluorescence signal. In this way, it was even possible to obtain X-ray absorption spectra (XAFS) of dilute elements in melts and solutions at high P and T (e.g., Wilke et al. 2006, Mayanovic et al. 2007). A substantial contribution to the background of the fluorescence spectra stems from inelastic scattering of the incoming beam as it travels through the

upstream diamond anvil. Acquisition of the XRF spectra with the detector at 90° to the incoming beam minimizes the inelastic scattering signal for linearly polarized synchrotron radiation because the scattering cross-section along the E vector is zero (e.g. Haller & Knöchel 1996, Schmidt & Rickers 2003). However, a substantial amount of elastically and inelastically scattered radiation is recorded even in this configuration because the detector's solid angle is finite and not only signal along the E vector is recorded. Furthermore, the focused beam is blurred by a halo of scattered radiation produced in the diamond anvil, which deteriorates the spatial resolution of the setup. This effect may limit the applicability of the technique for samples that are heterogeneous on the microscopic scale. A high spatial resolution is also important for DAC experiments at extreme pressures (up to 100 GPa), in which the total sample size is only in the range of 50 μm.

Here, we tested a confocal micro-XRF setup to overcome these limitations. The confocal setup is similar to the one proposed by Kanngiesser et al. (2003), Janssens et al. (2004) and Vincze et al. (2004). In these experiments, a probing volume is defined by coinciding the focal spot of an additional X-ray optic in front of the detector with the focal spot of the incoming beam. This arrangement minimizes signal contributions from the space outside the focal spot of the beam and even allows acquisition of chemical information in 3D (e.g., Kanngiesser et al. 2003, Janssens et al. 2004, Vincze et al. 2004). While the normal confocal setup is optimized for spatial resolution, a confocal setup for a DAC has to find a trade-off between spatial resolution and the focal distance that is required by the DAC. In the setup presented here, we used a glass polycapillary with a long focal distance as X-ray half-lens in front of the detector.

Experimental

Detector polycapillary

The polycapillary half-lens was designed to provide a focal spot of ca. 150-300 μm at a focal distance of 50 mm. It was optimized to work between 6 and 18 keV to cover a wide range of fluorescence lines accessible in DAC experiments. At lower energies, absorption of fluorescence X-rays by the diamond and the air between detector and DAC becomes dominant. At higher energies, the efficiency of the polycapillary deteriorates due to the critical angle for total reflection and the significant transmission of the hard X-rays through the capillary material. Additionally, the capillary was designed to cover the solid angle predetermined by fluorescence acquisition in 90° geometry for the DACs used here. Design

and manufacturing of the polycapillary were done by XOS©, East Greenbush, NY (USA). The detector polycapillary tube has a length of 65 mm and a diameter of 12 mm. The optical input diameter is 4.85 mm and the output diameter 7.57 mm. Entrance and exit are covered by a Be foil of 12.7 μm thickness. Transmission of the polycapillary is 23.4 % at 17.4 keV and 46.7 % at 8 keV as measured by the manufacturer using X-ray tube sources. The polycapillary was characterized further by knife-edge scans of thin Au and Nb foils using the synchrotron radiation beam at beamline L, HASYLAB.

The polycapillary is mounted on the nozzle of the fluorescence detector using an adapter made of aluminum (Fig. 1). The position of the detector with the mounted polycapillary has to be aligned to the focal spot of the incoming beam. This alignment is achieved using the signal of a thin metal foil positioned in the focal spot of the incoming beam. The foil is oriented at 45° to the beam to permit acquisition of the fluorescence signal at 90°. The position of the detector capillary is optimized by scans of the detector position in the vertical direction and in the direction parallel to the beam. The nominal distance of 50 mm between capillary and focal spot was verified by measuring the focal spot with the distance set to values below and above this nominal distance. At each distance, the size of the probed spot is determined by scanning a thin metal foil through the probing volume defined by focus of the beam and the detector capillary (see also Schmitz et al. 2009). The scan is done in the direction perpendicular to the foil surface, i.e., at 45° to the incoming beam. Such scans were also used to characterize the spatial resolution of the confocal setup. For these tests, Cu and Nb foils were used to measure the effect of fluorescence energy on the spatial resolution.

Beamlines

The setup was tested at two beamlines: beamline L, HASYLAB and ID24, ESRF. Beamline L uses synchrotron radiation from a bending magnet at the DORIS III storage ring that is run with positrons at an energy of 4.5 GeV and 80-140 mA ring current. A large-bandwidth multilayer monochromator was used to achieve a monochromatic beam with high photon flux. The multilayer monochromator consists of a Ni/C structure. The monochromator was adjusted to an excitation energy of 20 keV. The beam was focused using a single-bounce capillary that allows focusing of X-rays up to 40 keV down to a spot size of ca. 10 μm (Falkenberg et al. 2003, Schmidt et al. 2007) with a focal distance of 50 mm (see also Fig. 1). An ionization chamber filled with air was used to measure the intensity of the incoming beam.

The fluorescence was recorded using an energy-dispersive Vortex© Si drift-chamber solid-state detector.

Beamline ID 24 is located at an undulator source of the ESRF, where the storage ring is run with electrons at an energy of 6 GeV and 200 mA ring current. The optical layout at ID 24 is designed for dispersive X-ray absorption spectroscopy (XAFS) (Pascarelli et al. 2006). In the so-called TURBO-XAS mode, XAFS spectra can be acquired using the fluorescence yield for samples with low concentrations of the element of interest (Pascarelli et al. 1999). In this mode, a narrow slit is scanned through the energy dispersed radiation fan downstream of the polychromator to record the fluorescence yield as a function of the X-ray energy. For the experiment here, we used a Si(111) Bragg polychromator to achieve the energy resolution necessary to perform XAFS measurements. The optical layout consists in two Si mirrors in a Kirkpatrick Baez configuration, followed by the crystal polychromator and by a third vertically refocusing mirror positioned just before the sample. Further details of the optical layout are reported in Pascarelli et al. (2006). The diameter of the spot was 7 μm FWHM at ~10 keV. The intensity of the incoming beam was measured with a Si photo diode located between the last mirror and the DAC, with the diode recording radiation scattered from a Kapton foil. The fluorescence was recorded using an energy-dispersive Vortex© Si drift-chamber solid-state detector (see also Stechern et al. 2009).

Hydrothermal diamond-anvil cell

A hydrothermal diamond-anvil cell (HDAC) optimized for XRF measurements was used for the experiments (Schmidt & Rickers 2003, Schmidt et al. 2007, Borchert et al. 2009). A sketch of the HDAC is shown in Fig. 2. The sample chamber of the cell consists of a cylindrical hole in a polished Re or Ir gasket and a recess in the culet face of one diamond anvil. The gasket seals the sample chamber and separates the two diamond anvils. The gaskets used here had an initial diameter of ca. 500 μm and a thickness of 120 μm. The recess in the diamond anvil has a diameter of 200 μm and a depth of 60 to 80 μm. The recess allows acquisition of the fluorescence signal at an angle of 90° to the incident beam, which minimizes the background by scattered radiation. Additionally, the recess represents most of the sample volume from which fluorescence X-rays are collected. The solid angle accessible for fluorescence detection is about 0.2 x 0.3 rad, and is defined by the gasket, an opening in the cement with which the diamonds are mounted, and the window in the cover ring of the cell. The HDAC is heated by NiCr coils around the tungsten-carbide seats. Two K-type

thermocouples attached to the diamonds were used to measure the temperature. At room temperature, the assemblage in the sample chamber consisted of solid starting materials, an aqueous liquid (e.g. H_2O or NaCl-H_2O solution) and a vapor bubble. The actual electrolyte concentration of the solution in the sample chamber was obtained from cryometry, i.e., measurement of the vapor-saturated liquidus temperature. The density of the fluid was determined from the temperature of vapor-liquid homogenization. The pressure at a given temperature, concentration, and density was calculated using appropriate equations of state or correlation functions (Schmidt & Rickers 2003, Schmidt et al. 2007, Borchert et al. 2009). The sample chamber was monitored optically using a microscope to record all phase transitions and sample changes that occur during freezing, heating, annealing and quenching. Element concentrations in the aqueous fluid at P and T were determined by scaling the fluorescence peak net intensity of an unknown with the net intensity collected for this element on a standard solution in the sample chamber of the cell. Spectra of standard solution and unknown were acquired with the same excitation conditions and geometrical setup and both net intensities are normalized to the density of each solution. This direct calculation of the concentration in the unknown is valid if the concentrations of major components in both sample and standard solution are similar (e.g., Schmidt & Rickers 2003, Schmidt et al. 2007). If the solution composition for the unknown differs significantly from that of the standard solution, a correction was applied for the absorption of the fluorescence along the path through the fluid in the recess (see also Manning et al. 2008).

Results & Discussion

The detector polycapillary was tested at beamline L, HASYLAB, with an energy of the incident beam of 20 keV. At this energy, the transmission of the polycapillary was 19.1 % using a horizontal beam dimension of 3 mm and a vertical dimension of 1.5 mm at the entrance slit. This result is consistent with the values determined by the manufacturer at 17.4 keV (23.4 %), because the transmission decreases with increasing X-ray energy. An edge scan on a Nb foil with a thickness of 2.5 μm revealed a beam size of 148 µm (FWHM) at 20 keV. This measured value agrees well with the design parameters given by the manufacturer (≤ 160 μm at 17.4 keV and ≤ 300 μm at 8.0 keV), which are based on simulation data. A gain factor of 50 at 16.6 keV was obtained from comparison of the flux through the entrance slit (1.5 x 3 mm^2) with that transmitted through the polycapillary.

The spatial resolution defined by the aligned capillaries in the incoming beam and in front of the detector was measured by scanning the probing volume with thin metal foils. Figure 3 shows the fluorescence intensity as a function of the stage position of one scan using an 8 μm thick Cu foil and one scan on a 2.5 μm thick Nb foil. Both scans were done at beamline L, HASYLAB. The FWHM of the peaks are 225 μm in the case of the Cu Foil (8 keV) and 108 μm for the Nb foil (16.6 keV). The FWHM of the measured profiles can be related to the probing volume and the thickness d of the foil (Janssens et al. 2004)

$$FWHM^2_{profile} = FWHM^2_{SBC} + FWHM^2_{DPC} + d^2_{foil},$$

where $FWHM^2_{SBC} + FWHM^2_{DPC}$ describes the width of the probing volume defined by the single-bounce capillary and the detector polycapillary. Using this equation and the known values for the single-bounce capillary and the foil thickness, the width of the focal spot of the detector polycapillary was calculated from the measured profiles. This resulted in the same values as those determined directly on the profiles, i.e. 108 μm and 225 μm. Thus, in this type of scan, which characterizes the depth of the probing volume in the direction 45° to the incoming beam, the detector capillary governs the width of the profile.

Additional information on the spatial resolution at the conditions of in-situ experiments is provided by scans across HDAC sample chambers that contained zircon ($ZrSiO_4$) or hafnon ($HfSiO_4$) crystals. Such a sample chamber is shown in Fig. 4a with a zircon crystal located at the lower rim of the gasket and aqueous solution, in which a portion of the zircon had dissolved. For this measurement the cell was rotated by 3° around the vertical axis in respect to the beam, which maximizes the solid angle for the fluorescence in the direction of the detector and avoids reduction of the signal by absorption in the gasket. In a vertical scan across this chamber (Fig. 4b), the Zr K_α signal from the solution in the recess can be well separated from that from the zircon crystal. Therefore, the position of the crystal in the sample chamber relative to the recess is not as crucial as in experiments without detector capillary, where care must be taken that the signal from the solution in the recess is not superimposed by signal from co-excitation of the crystal by scattered X-rays of the incident beam. E.g., in the study of Schmidt & Rickers (2003) the crystal was placed in an extra pocket drilled into the rim of the gasket to avoid co-excitation.
Figure 4c shows a similar example, where the cell was not rotated, i.e. in line with the axis of the beam. In this case, the vertical scan across the chamber (Fig. 4d) shows only Zr K_α signal from the recess, whereas fluorescence from the zircon crystal is not visible at all. In this

orientation of the cell, the confocal setup thus allows a complete suppression of the signal by the crystal at any location in the gasket hole. In Fig. 4e an example is shown, where a hafnon crystal had shifted closer to the recess during the run and a second Hf-bearing crystal had grown at the lower rim of the recess. In the first scan of the Hf-L_α intensity across the sample chamber, the hafnon crystal at the margin of the sample chamber is clearly visible from the sharp increase in intensity. However, this signal does not affect the intensity representative for the Hf concentration in the fluid measured in the recess. The second scan shows that the crystal shifted towards the recess, and that a second Hf-bearing crystal grew at the rim of the recess. However, the transition of the signal from the crystal to remaining recess is still very sharp. More importantly, the Hf-L_α intensity in the remaining portion of the recess is very close to the one in the first scan. Both scans indicate that determined Hf concentrations for the fluid based on the measured L_α intensity in the recess are not affected by co-excitation of the solid Hf phases even under this unfavorable geometric arrangement of crystal and recess.

Figure 5a compares XRF spectra measured at beamline L, HASYLAB, on a 1300 ppm Zr standard solution in the sample chamber of a HDAC. One spectrum was acquired using the confocal setup with the detector capillary, whereas the other one was measured using a collimator on the fluorescence detector with 2 mm diameter at a distance of 50 mm from the center of the HDAC. First of all, the total signal recorded by the detector when using the detector capillary is drastically decreased. This decrease is due to a smaller solid angle seen by the detector, the transmission of the polycapillary, which is smaller than 100%, and higher absorption because of the longer distance between probing volume and detector. For Zr K_α, the intensity is reduced by a factor of 3.4 for the spectrum acquired with the capillary. However, the ratio between Zr K_α peak maximum and the maximum of the Compton scattering increases from 0.5 in the spectrum recorded with the collimator to 1.3 if the detector capillary was used. Inspection of the spectrum acquired with the collimator indicates that the influence of the Compton scattering to the measured signal extends down to about 14 keV at an excitation energy of 20 keV. The spectrum acquired with the detector polycapillary indicates a much lower intensity of the scattered radiation (by factor of 10). The contribution to the signal becomes already negligible between 15 and 16 keV. The strong signal from elastic and inelastic scattering is produced along the path of the incident beam through the diamond anvil. The detector polycapillary efficiently suppresses collection of this signal. In addition, the intensity of Re fluorescence lines from the gasket are decreased by a factor of 12. Finally, the spectrum collected with the collimator shows a Sr K_α peak at 14.2 keV, which

is not present in the other spectrum and thus indicates a contribution generated outside the sample chamber.

The spectra shown in Fig. 5b were taken on a 1000 ppm Hf standard solution and on a NaOH-solution which contained ca. 400 ppm Hf due to equilibration with a hafnon crystal at 400°C. Both spectra were acquired at ID 24, ESRF. Fluorescence peaks of Fe, Ni and Zn are present in addition to the L-lines of Hf. The intensities of the Fe, Ni, and Zn lines correlate with those of the Hf-lines. The exact source for these lines could not be determined, but they are probably generated by secondary excitation of these elements along the path of the fluorescence and the scattered radiation to the detector.

Detection limits (DL) for this setup were estimated based on the spectra measured on several standard solutions from the relationship

$$DL = C_{STD} * 3 * sqrt(I_{BC})/I_{Peak}$$

where C_{STD} denotes the concentration of the standard solution, I_{BC} the intensity of the background, and I_{Peak} the intensity of the fluorescence peak (e.g., Haller & Knöchel 1996). Peak and background intensities were determined using the software package AXIL (e.g., Vekemans et al. 1995). Using the K_{α}-peak, a relative DL of 1 ppm was obtained for Zr (500 s acquisition time) for the setup with detector polycapillary at beamline L. This value is similar or even slightly better than that obtained with the classical setup. The DL for the classical setup range between 1-3 ppm, including earlier measurements of Schmidt et al. (2006) for the HDAC setup at beamline L (1000 s acquisition time). At the used excitation conditions, the fluorescence spectra taken at ID 24 yield a DL for Hf of 0.5 ppm based only on the L_{α}-peak (500 s acquisition time). Both cases show that the confocal setup leads to sensitivities that are similar or even better than those obtained with the classical setup. Enhancement of the DL certainly stems from the improved peak-to-background ratio. However, the enhancement is limited by two other parameters: (i) the transmission of the polycapillary is smaller than 100% (see Experimental section); (ii) the distance between the detector and probing volume is significantly longer, so that absorption reduces the fluorescence intensity particularly at lower energies (e.g. in case of Hf). The enhancement of DL that was observed here is consistent with earlier results by Janssens et al. (2004) for the confocal setup, who report improvements especially for thick samples, where background from inelastic scattering is more severe.

Finally, the confocal setup was also applied at ID 24 to record fluorescence XAFS spectra at the Hf L-edge on aqueous solutions at high temperature and pressure. Two examples are shown in Fig. 6, which illustrate the potential for acquisition of XANES or EXAFS spectra at the given concentrations. At a solute concentration of ~400 ppm Hf, the signal was weak, so that it was only worthwhile to acquire an extended XANES range, with 40 s/pt. At 5000 ppm Hf, acquisition of EXAFS spectra became feasible, as demonstrated by the spectrum of Hf in the $Na_2Si_3O_7$-bearing aqueous solution, collected with 50 s/pt. The spectrum corresponds to an EXAFS range of 8 $Å^{-1}$. It is noted here that all XAFS spectra collected in this study showed very little distortions in the raw data, i.e., fluorescence yield normalized to the intensity of the incoming beam. This does not only indicate an appropriate setup for measurement of the intensity of the incoming beam, but also that the confocal setup reduces contributions from scattered radiation to the background of the fluorescence lines of interest (here Hf). Usually, such a background contribution is excitation-energy dependent because of the increasing difference between fluorescence and excitation energy, which changes the background and leads to tilting or distortion of the XAFS spectrum.

Conclusion & Outlook

The results shown here demonstrate the advantages of a confocal micro-XRF setup for experiments with diamond anvil cells. This setup enhances the quality of the fluorescence spectra, and thus the sensitivity for detecting elements at low concentrations. The confocal X-ray optical setup significantly facilitates HDAC experiments, particularly those involving the dissolution of substances in fluids. This is because it efficiently suppresses unwanted fluorescence signal from inside and outside the sample chamber of the DAC as well as contributions by elastically and inelastically scattered radiation of the incoming beam formed on the path through the diamond anvil. Specifically, the actual location of a mineral relative to the recess in the sample chamber of the HDAC is less crucial for confocal spectrum acquisition, because it is easier to avoid collection of XRF from co-excitation of the mineral by scattered X-rays in the upstream diamond anvil. The examples shown were acquired with detection at 90° to the incoming synchrotron radiation beam. However, most DACs do not allow the detection of the fluorescence signal at this angle, or only if a Be gasket is used, which is transparent to X-rays, but poses many other experimental limitations. Sanchez-Valle et al. (2003) used a membrane-driven DAC and a position of the fluorescence detector at 15° to the incoming beam. In this forward-scattering geometry, however, the signal from Compton scattering becomes huge. A confocal setup similar to the one proposed here should

provide an efficient means for reducing this signal, and would thus greatly enhance the sensitivity of XRF analyses in forward-scattering DAC experiments.

Acknowledgements

We thank M. Kreplin, R. Schulz, the staff of the high-pressure workshop at GFZ, W. Ternes at DESY as well as F. Perrin at ESRF for technical help and support. Financial support by the DFG through the grant Wi 2000/5-1 and by the GFZ through W. Heinrich is highly appreciated. HASYLAB at DESY and the ESRF are thanked for granting beam time.

References

Boehler R. (2005). In: Mineral Behaviour at Extreme Conditions, Ed. R. Miletich, *EMU Notes in Mineralogy* **7**, 217-224.

Borchert, M., Wilke, M., Schmidt, C., Rickers, K. (2009). *Chem. Geol.* **259**, 39-47.

Falkenberg, G., Rickers, K., Bilderback, D.H., Huang, R. (2003). *HASYLAB Annual Report*, 71–74.

Haller M. , Knöchel A. (1996). *J. Trace and Mircroprobe Techniques* **14**, 461-488.

Janssens, K., Proost, K., Falkenberg, G. (2004). *Spectrochim. Acta B* **59**, 1637-1645.

Kanngiesser, B., Malzer, W., Reiche, I. (2003). *Nucl. Instrum. Methods B* **211**, 259-264.

Keppler H., Frost D.J. (2005). In: Mineral Behaviour at Extreme Conditions, Ed. R. Miletich, *EMU Notes in Mineralogy* **7**, 1-30.

Manning, C.E., Wilke, M., Schmidt, C., Cauzid J. (2008). *Earth Planet. Sci. Lett.* **272**, 730-737.

Mayanovic R., Anderson A.J., Bassett W.A., Chou I.M. (2007). *Rev. Sci. Instr.* **78**, 053904.

Pascarelli S, Neisius T, De Panfilis S (1999). *J. Synchrotron Rad.* **6** 1044-1050.

Pascarelli S, Mathon O, Munoz M, Mairs T, Susini J (2006). *J. Synchrotron Rad.* **13** 351.

Sanchez-Valle C., Martinez I., Daniel I., Philippot P., Bohic S., Simionovici A. (2003). *Am. Mineral.* **88**, 978-985.

Sanchez-Valle C., Daniel I., Martinez I., Simionovici A., Reynard, B. (2004). *J. Phys. Condens. Matter* **16**, S1197-S1206.

Schmidt, C., Rickers, K. (2003). *Am. Mineral.* **88**, 288-292.

Schmidt, C., Rickers, K., Wirth, R., Nasdala, L., Hanchar, J. (2006). *Am. Mineral.* **91**, 1211-1215.

Schmidt, C., Rickers, K., Bilderback, D.H., and Huang, R. (2007). *Lithos* **95**, 87-102.

Schmitz, S., Möller, A., Wilke, M., Malzer, W., Kanngiesser, B., Bousquet, R., Berger, A., Schefer, S. (2009). *Eur. J. Mineral* **21**, 927-945.

Stechern, A., Wilke, M., Schmidt, C., Rickers, K., Pascarelli, S., Manning, C.E. (2009)- *J. Phys. Conf. Ser.* **190**, 012058.

Vekemans B., Janssens K., Vincze L., Adams F., Van Espen P. (1995). *Spectrochim. Acta Part B* **50**, 149-169.

Vincze, L., Vekemans, B., Brenker, F.E., Falkenberg, G., Rickers, K., Somogyi, A., Kersten, M., Adams, F. (2004). *Anal. Chem.* **76**, 6786-6791.

Wilke M., Schmidt C., Farges F., Malavergne V., Gautron L., Simionovici A., Hahn M., Petit P.E. (2006). *Chem. Geol.* **229**, 144-161.

Figure Captions

Figure 1: Photograph showing a top view of the experimental setup at beamline L. The HDAC is mounted on a motorized XYZ sample stage and can be rotated around the vertical axis. The confocal probing volume defined by the single-bounce capillary and the detector capillary is positioned at the center of the cell.

Figure 2: Schematic drawing showing the details of the hydrothermal diamond-anvil cell designed for the in-situ analysis of fluids at high pressures and temperatures using XRF analysis. Modified after Schmidt et al. (2007).

Figure 3: Fluorescence intensity vs. stage position for scans of a thin Cu-foil (a) and a thin Nb-foil (b) through the probing volume as indicated. Full-width-at-half-maximum (FWHM) values given were determined from the data.

Figure 4a: Photograph of the sample chamber of the HDAC containing a zircon crystal ($ZrSiO_4$) and an aqueous solution at 750°C.

Figure 4b: Vertical scan of Zr-K_α intensity normalized to the incident beam intensity across the sample chamber shown in Fig. 4a. Error bars indicate standard deviation of measured signal. Measurement was performed at beamline L.

Figure 4c: Photograph of the sample chamber of the HDAC containing a zircon crystal ($ZrSiO_4$) and an aqueous solution at 600°C.

Figure 4d: Vertical scan of Zr-K_α intensity normalized to the incident beam intensity across the sample chamber shown in Fig. 4c. Error bars indicate standard deviation of measured signal. Measurement was performed at beamline L.

Figure 4e: Vertical scans of the Hf-L_α intensity normalized to the incident beam intensity across the sample chamber containing a hafnon crystal ($HfSiO_4$) at various positions to the recess. See text for details. Scans show two points in time. Error bars indicate standard deviation of measured signal. Measurements were performed at ID 24.

Figure 5a: X-ray fluorescence spectra of a standard solution containing 1300 ppm Zr loaded into the HDAC. The spectra were measured at beamline L with the detector capillary or detector collimator as indicated.

Figure 5b: X-ray fluorescence spectra of a standard solution containing 1000 ppm Hf loaded into the HDAC measured at room temperature and of a NaOH solution (35 wt%) equilibrated with a hafnon crystal at 500°C (ca. 400 ppm Hf). The spectra were measured at ID 24 using the detector capillary.

Figure 6: Fluorescence XAFS spectra taken at ID 24 on solutions in the HDAC at the conditions indicated.

Figures

Fig. 1

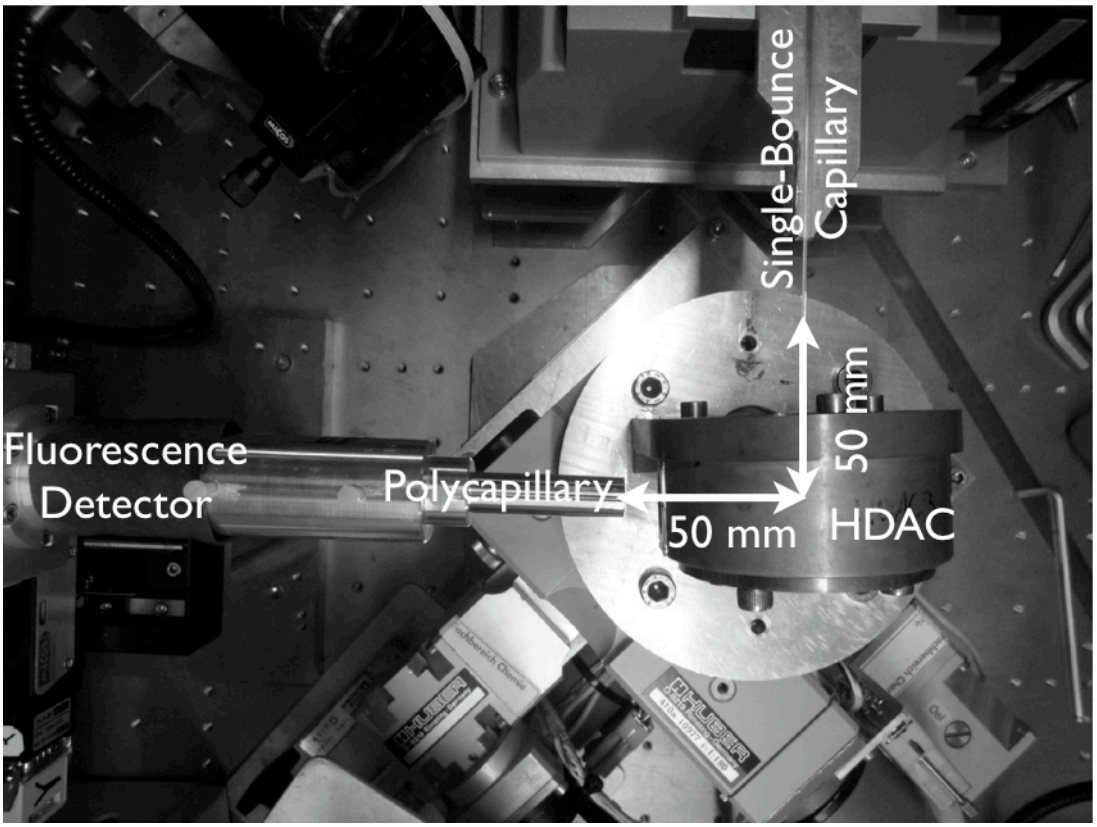


Fig. 2

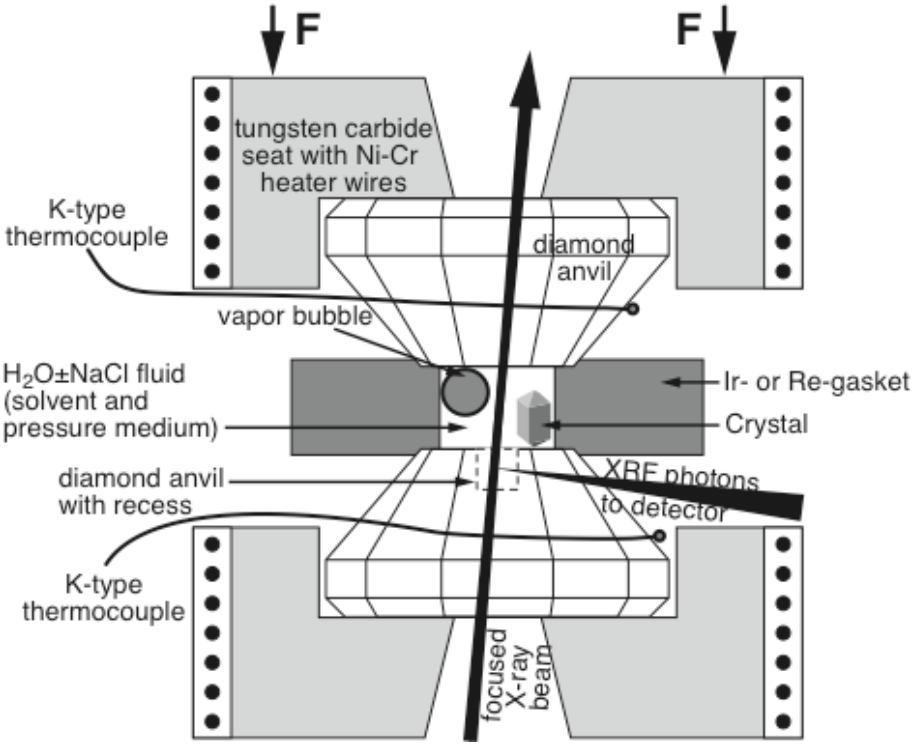


Fig. 3a

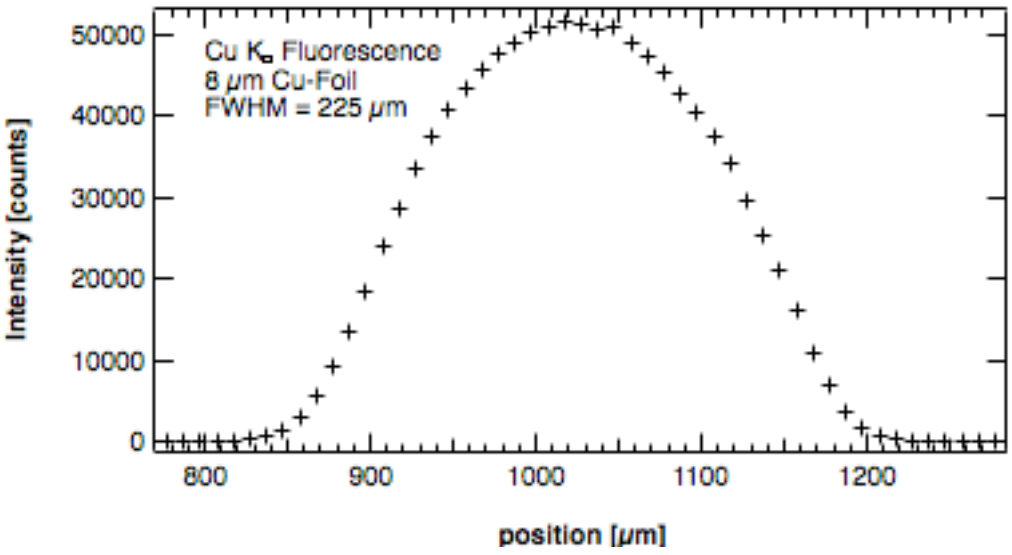

Cu K$_\alpha$ Fluorescence
8 μm Cu-Foil
FWHM = 225 μm
Intensity [counts]
position [μm]
0
10000
20000
30000
40000
50000
800
900
1000
1100
1200

Fig. 3b

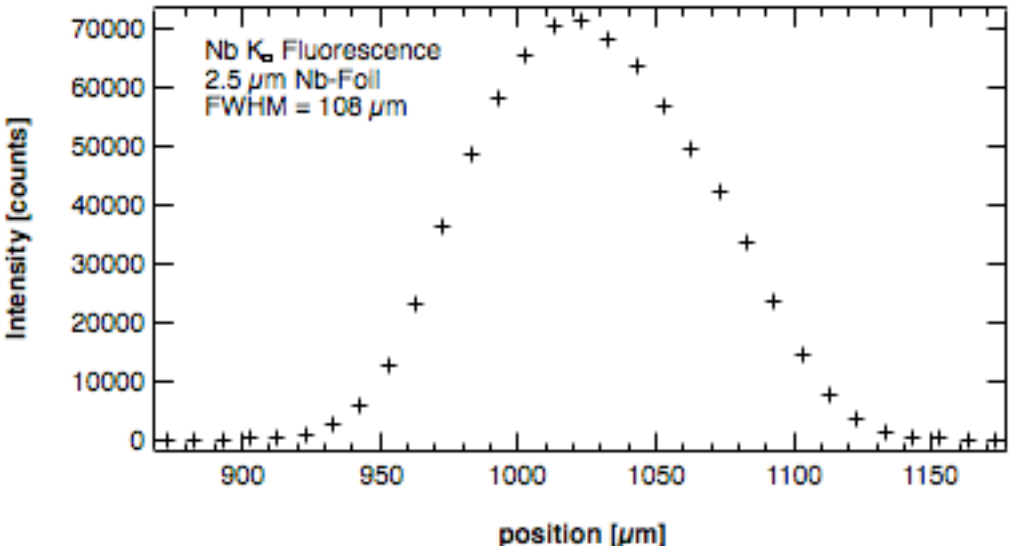

Nb K$_\alpha$ Fluorescence
2.5 μm Nb-Foil
FWHM = 108 μm
Intensity [counts]
position [μm]
0
10000
20000
30000
40000
50000
60000
70000
900
950
1000
1050
1100
1150

Fig. 4 a)

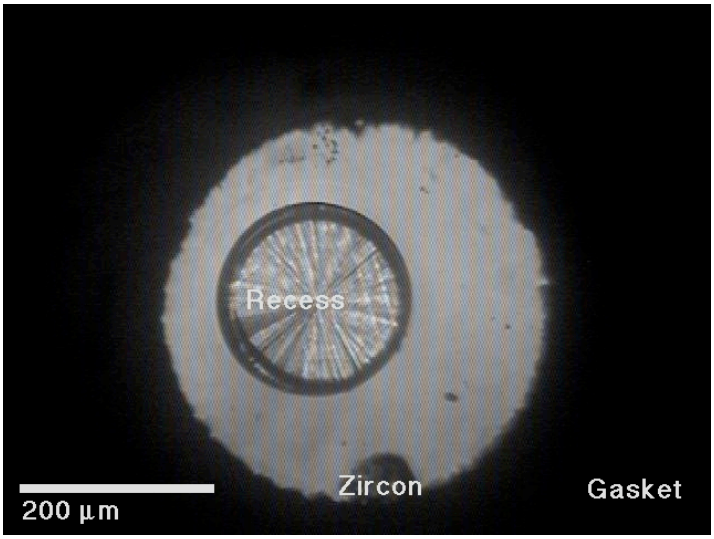

Recess
Zircon
Gasket
200 µm

Fig. 4b)

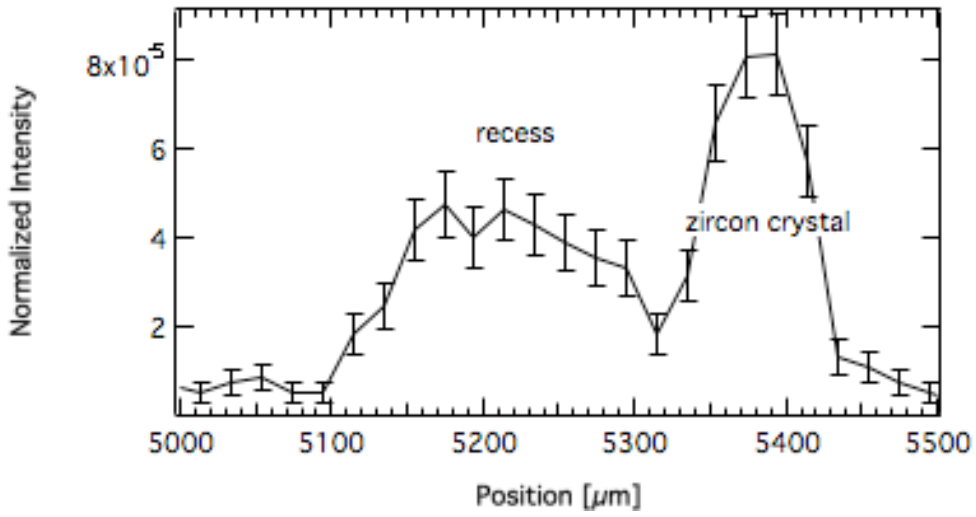

Normalized Intensity
8x10^5
6
4
2
recess
zircon crystal
5000
5100
5200
5300
5400
5500
Position [µm]

Fig. 4c)

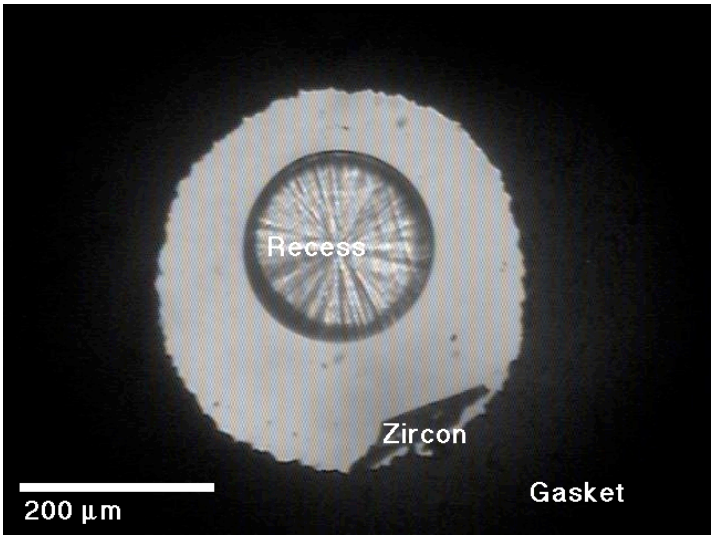


Fig. 4d)

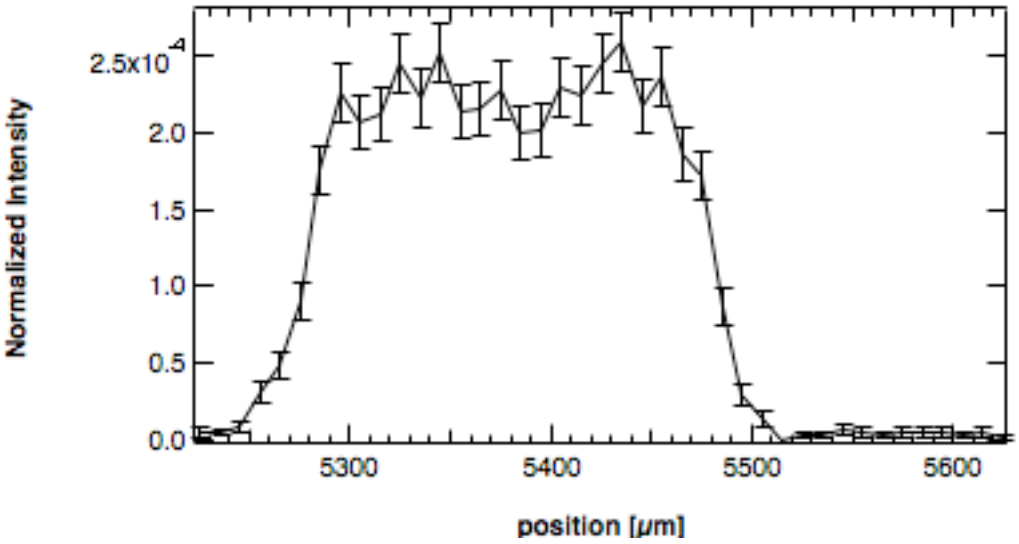


Fig. 4e)

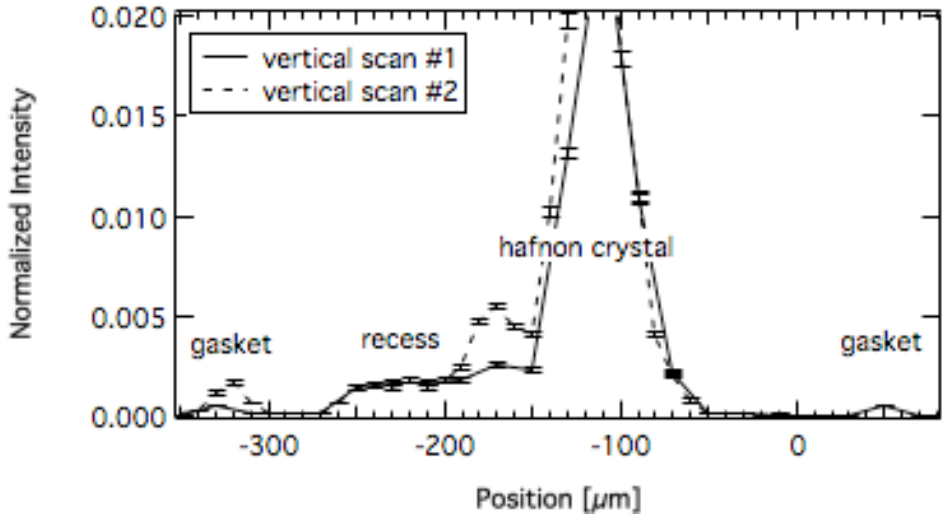


Fig. 5a)

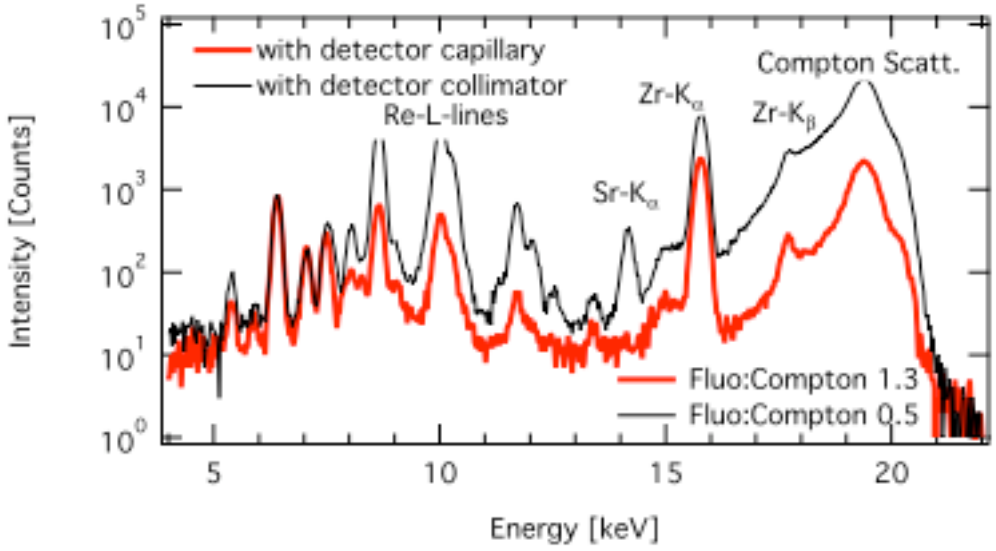

with detector capillary
with detector collimator
Re-L-lines
Zr-Kα
Zr-Kβ
Compton Scatt.
Sr-Kα
Fluo:Compton 1.3
Fluo:Compton 0.5
Intensity [Counts]
Energy [keV]

Fig. 5b)

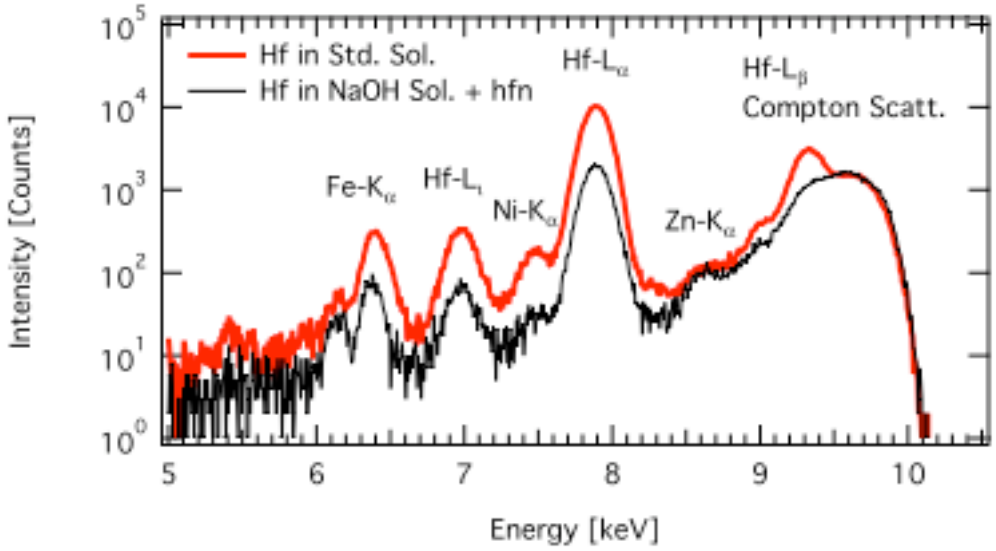
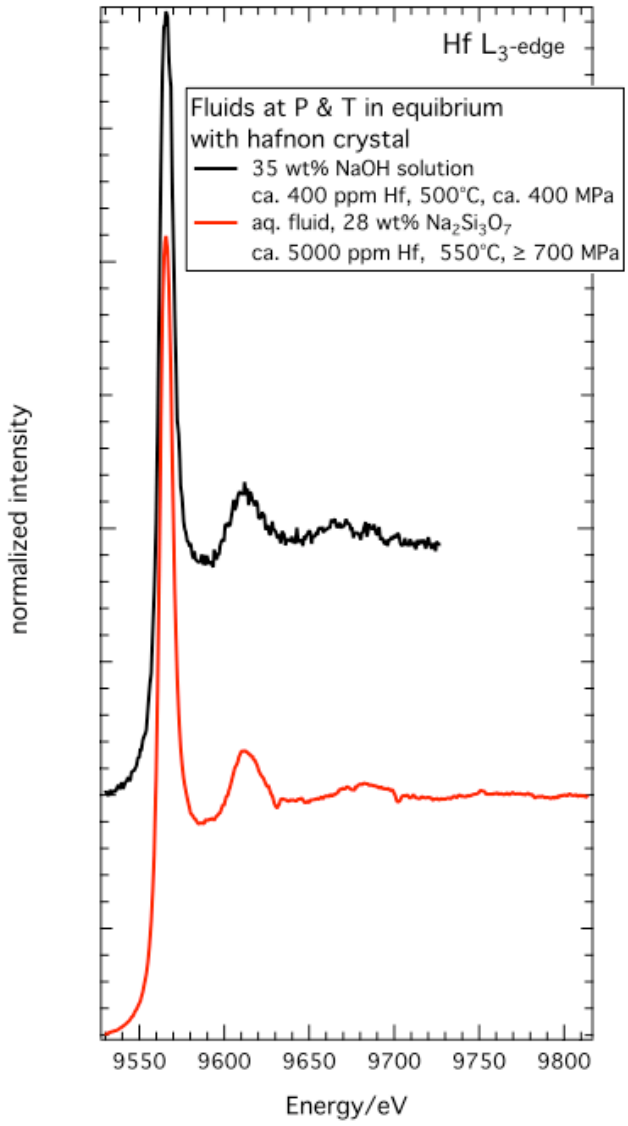

Hf in Std. Sol.
Hf in NaOH Sol. + hfn
Hf-Lα
Hf-Lβ
Compton Scatt.
Fe-Kα
Hf-Lι
Ni-Kα
Zn-Kα
Intensity [Counts]
Energy [keV]

Fig. 6


Hf L3-edge
Fluids at P & T in equibrium
with hafnon crystal
35 wt% NaOH solution
ca. 400 ppm Hf, 500°C, ca. 400 MPa
aq. fluid, 28 wt% Na2Si3O7
ca. 5000 ppm Hf, 550°C, ≥ 700 MPa
normalized intensity
9550
9600
9650
9700
9750
9800
Energy/eV